

You have 80 minutes for this exam.

October 7, 2008

Exams written in pencil or erasable ink will not be re-graded under any circumstances.

Explanations should be concise and clear. I have given you more space than you should need.

N = 74

You need a calculator for this exam, and no other study aids or materials are permitted.

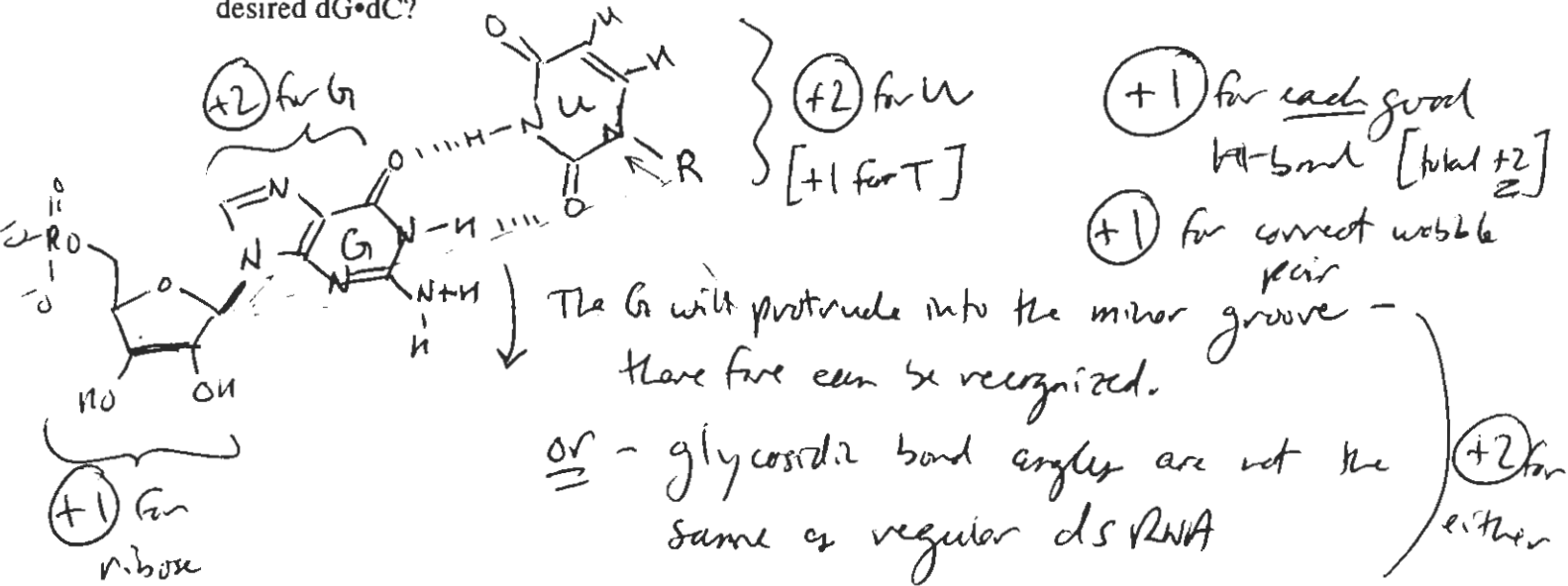
Generous partial credit will be given, i.e., if you don't know, guess.

Honor Pledge: At the end of the exam time, please write out the following sentence and sign it, or talk to me about it:

"I pledge on my honor that I have not given or received any unauthorized assistance on this examination."

1. DNA Structure and Base Pairing (30 pts):

(a; 10 pts) Draw a rG•rU wobble pair in RNA, showing the sugar for one of the nucleotides. Given that the related dG•dT base pair is quite stable, how do you think biology distinguishes it from the desired dG•dC?



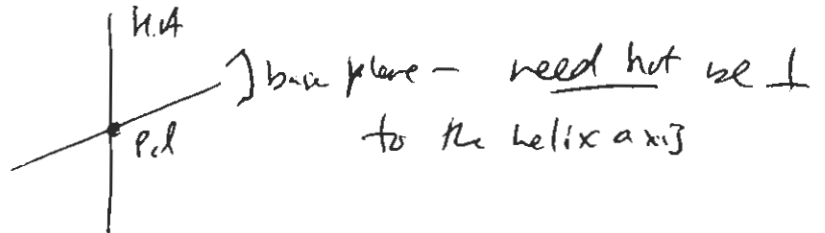
(b; 10 pts) We discussed DNA symmetries. Each base pair has a pseudodyad axis (rotation by 180° about a dyad axis means rotates an object onto itself). What is "pseudo" about the base pair pseudodyad - what part of the base pair is rotated onto itself? Explain why the helix axis is necessarily perpendicular to the pseudodyad. Circle Yes or No to answer whether the helix axis necessarily perpendicular to the base pair plane.



relates glycosidic bonds (+3)

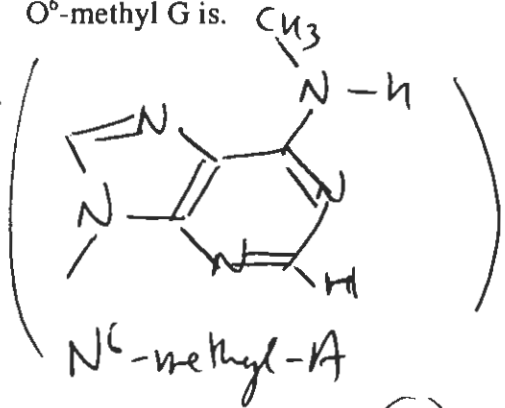
not the complete bp or structure (+3)

(+3) { If the H.A. were not \perp to the p.d., then it would not be rotated into itself.

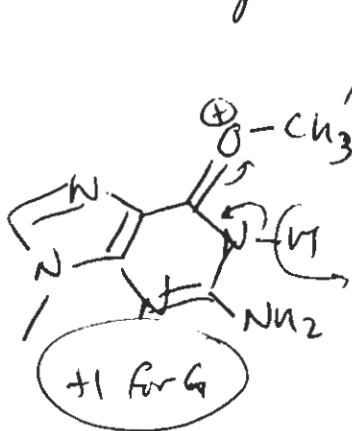


(c; 10 pts) Draw the structures of N⁶-methyl A (i.e. an A with a methyl group attached to the amino group at position 6) and O⁶-methyl G. Explain why N⁶-methyl-A is not a miscoding lesion, whereas O⁶-methyl G is.

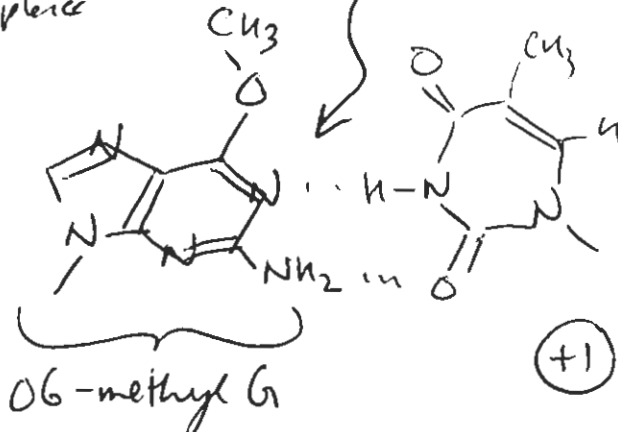
(+1) for A
(+1) for -CH₃ on N⁶



Same W-C face (+1)



(+1) for methyl in the right place



(+2) for recognition of tautomerization

(+2) W-C H-bonding face is different - what was a donor is now an acceptor - can H-bond to T instead of C

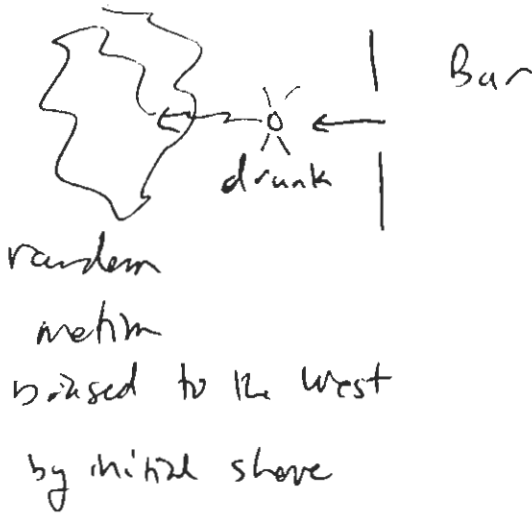
2. Secondary and tertiary structure (20 pts):

(a; 8 pts) What are the two most important structural elements stabilizing tertiary structure in RNA? How could you destroy tertiary structure in an RNA without disrupting secondary structure?

(+2) - Non-Watson-Crick hydrogen bonds
 Divalent \leftarrow (+1)
 - Metals chelating phosphates that approach each other
 (+1) \leftarrow (+1)

(+3) - Either: chelate away all divalent metals M^{++}
 or
 thermal denaturation [may not work in real life]

(b; 12 pts) We gave the analogy of the DNA trajectory as the path of a drunk being forcefully ejected from a bar; for the purposes of the problem, assume that the bar's door faces due west. In this analogy, what corresponds to the behavior of short vs. long lengths of DNA? How would one measure the persistence length for the drunk-ejection process. [Do not try this experiment in college!]



Short DNA \rightarrow travels along the initial direction due to impetus from bouncer's foot - projection \approx contour length
 (+3)

Long DNA - random motion leading to no net average movement in any direction
 (+3)

Measure persistence length -

(+3) for repeated trials + taking average
 (Eject 100's of drunks with same initial velocity [or come back night after night], measure where they end up after a long time, average displacement to the west = P)

(+3) for displacement in initial direction

3. DNA topology (30 pts):

(a; 8 pts) Consider a plasmid of 3000 bp. Assume the helical repeat is 10.5 bp/turn. Calculate Lk_0 , the ideal (non-integral) relaxed linking number and Lk_m , the actual (integral) linking number closest to Lk_0 . Plasmid DNA as isolated from cells typically has $\sigma \equiv \Delta Lk / Lk_0 = (Lk - Lk_0) / Lk_0 = -0.06$. Calculate ΔLk for this plasmid. (The ΔLk will be approximately equal to the writhe, as DNA is torsionally stiff.)

$$Lk_0 = \frac{N}{bp} = \frac{3000 \text{ bp}}{10.5 \text{ bp/turn}} = 285.7 \quad (+1 \text{ for setting up calculation}) \quad (+2 \text{ for } \#)$$

$$Lk_m = 286 \quad (+2)$$

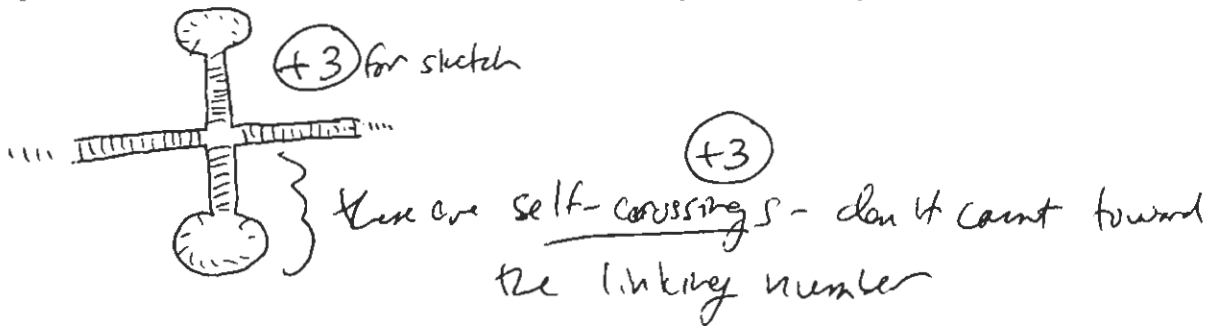
$$\sigma = \frac{\Delta Lk}{Lk_0} = -0.06 \quad \Delta Lk = -0.06 \times 285.7 = -17.1 \quad (+3)$$

[+1 if sign is wrong]

So the most common $\Delta Lk = -17$ - there will be ~~typo~~ topoisomers with $\Delta Lk = -15, -16, -17, -18, -19$

$wr \approx -17$ (actually more like -14, with the rest being ΔTw)

(b; 9 pts) Sketch a DNA cruciform structure. When a cruciform is extruded from a plasmid, the ~~linking~~ ^{twist} number is substantially reduced even though most of the DNA is still helical - why? In the example above, if 5 helical turns are extruded as a cruciform, and assuming that the ~~super~~ superhelical strain in the rest of the dsDNA is manifested exclusively as writhe, what is the value of Wr for the plasmid? (The decrease in writhe and hence bending is the driving force for cruciform extrusion.)



Extruding 5 helical turns $\rightarrow \Delta Tw = -5$, so $\Delta wr = -12$

(+3)

Biochemistry

(c; 9 pts) What are the two main differences between Type I and Type II topoisomerases?

Topoisomerase poisons are useful anti-cancer agents. What makes a topoisomerase halted in the middle of its catalytic cycle particularly toxic to a cell (e.g. as opposed to a random metabolic enzyme that is inactivated by a drug)?

Type I: Cuts + rejoins one strand, so $\Delta Lk = \pm 1$, does not burn ATP

Type II: Cuts + rejoins both strands, so $\Delta Lk = \pm 2$, does burn ATP

+3 for either/both - key on the same essentially [+4 if there are given as 2 reasons]

The topoisomerase is still linked to DNA - the protein itself becomes a massive DNA adduct that disrupts replication - it's not simply the loss of a needed activity.

+3

(Sideroidal)

(d; 4 pts) Draw a toroidal superhelix with $Wr = -4$.

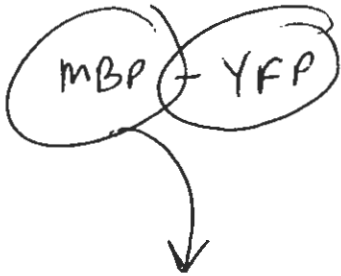
Left handed toroid -



+2 for a toroid
+1 for node signs
+1 for -4

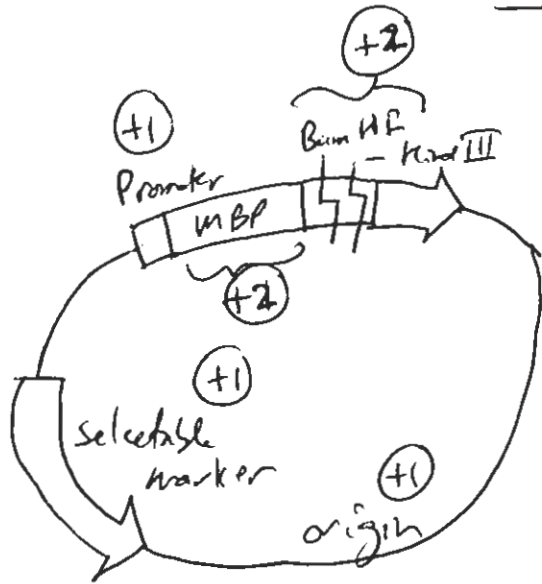
4. Methods (20 pts):

(a; 10 pts) Briefly describe the utility of making a fusion protein between MBP=maltose binding protein and YFP=your favorite protein. Assuming that you have a BamH I-Hind III fragment bearing the coding region of YFP, sketch a corresponding vector into which you would clone the fragment to make a fusion protein.



will stick to a maltose column - makes purification much easier (or immunofluorescence)

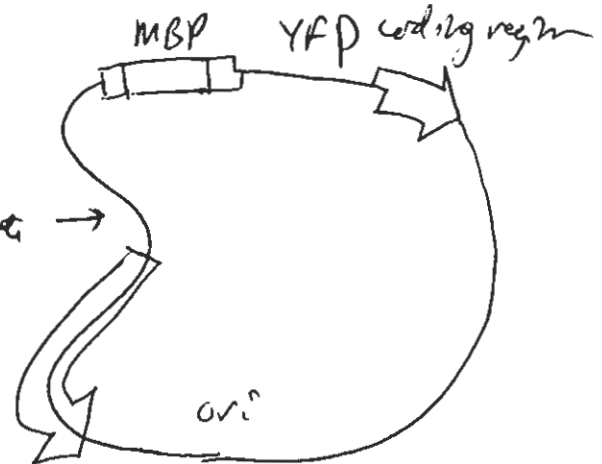
(+3)



B. YFP K

BamHI
K
HindIII

ligate →

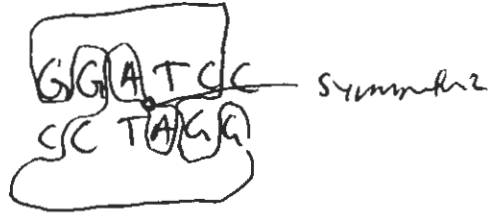


~~Handwritten scribble~~

(b; 10 pts) What is the natural function of restriction enzymes (REs)? Why do dimeric restriction enzymes typically have palindromic recognition sequences? Do REs require ATP hydrolysis to act? Why or why not?

(+2) -- They destroy foreign DNA that lacks the protective cognate methylation pattern

(+3) -- Each subunit makes identical interactions with the target sequence.



(+2) -- RE's do not require ATP hydrolysis

(+3) -- They are catalyzing a downhill reaction -- hydrolysis of a phosphodiester to a phosphomonoester

Page	Score
1	
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